

# Detection of *Mycoplasma hominis* and *Ureaplasma urealyticum* in Blood Samples of Recurrent Pregnancy Loss in Women by Polymerase Chain Reaction.

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**Summary:**

**Background:** Genital mycoplasma are implicated in pelvic inflammatory disease, puerperal infection, septic abortion, low birth weight, nongonococcal urethritis and prostatitis as well as spontaneous abortion and infertility in women.

**Objective:** We aimed to find a relationship between repeated abortions of unknown etiology and caused of *Mycoplasma hominis* and/or *Ureaplasma urealyticum*.

**Methods:** one hundred sixty cases, (15-49 years old) with history of recurrent abortion, intrauterine fetal death and/or neonatal death (after exclusion of other factors as cause abortion), and hundred women with normal pregnancy outcome with the same age were chosen as controls. *M. homini* and *U. urealyticum* were detected in blood by PCR.

**Result:** *M. homini* could be detected in 12\160 (7.5%) in women with pregnancy losses, but was not detected in control group. *U. urealyticum* could be detected in 64\160 (40%) in patient group and 4\100 (4%) in control group. The rapid detection of *M. homini* and/or *U. urealyticum* by PCR in pregnancy loss women could be important and necessary. The detection rate of *M. homini* and/or *U. urealyticum* in young women age (20-29 years) was higher than the others. Significant difference was observed in patients with three or four abortions compare with 2 or one abortion in addition to history of adverse outcome.

**Conclusion:** the role *M. homini* and/or *U. urealyticum* in the etiology of pregnancy losses was proposed.

**Key words:** *Mycoplasma hominis*, *ureaplasma urealyticum*, recurrent pregnancy loss, PCR.

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**Introduction:**

The etiologies of recurrent miscarriage are diverse and may be divided into genetic defects, such as chromosomal anomalies; maternal reproductive anatomic disease, both developmental and acquired, such as septate uterus or cervical incompetence and systemic maternal disease such as diabetes mellitus and infective factors. (1)Genital mycoplasmas represent a group of microorganisms that are commonly found in the genitourinary tract of pregnant and non-pregnant women. They have been associated with various pathological conditions and intrauterine infections, including pyelonephritis, pelvic inflammatory disease, chorioamnionitis, endometritis, and post partum fever, leading to important complications such as preterm birth, low birth weight, spontaneous abortion, still birth, premature birth, infertility, and perinatal mortality. (2,3,4,5)Infection with *Mycoplasma hominis* (*M.hominis*) or *Ureaplasma urealyticum* (*U.urealyticum*) two members of the class mollicutes are considered as the possible etiological agents in the family causing pregnancy loss since the frequently produce asymptomatic infection and are not identified by routine microbiological techniques. (1)Diagnosis of mycoplasma is usually made by serological determination or in vitro isolation of the organism. However, serological procedures are often

hampered by interspecies cross-reaction, while cultivation is time consuming and hard to for some fastidious mycoplasmas. Use of mycoplasma species- specific DNA probes made it possible to discriminate between different species, this method proved to be sensitive and specific. (6,7)The present study aimed at finding the relationship between fetal losses of unknown etiology and presence of mycoplasma infection (*Mycoplasma hominis* and *Ureaplasma urealyticum*) in blood samples from pregnancy losses by polymerase chain reaction (PCR) technique.

**Patients and methods:-**

**Subjects:-** two groups of women, age ranged between (15-49 years) were included in this study. The first group was 160 women with a history of pregnancy loss, such as, repeated abortion, intrauterine fetal death and/ or neonatal death. Medical examination and family history complete physical examination, laboratory investigations were done to exclude other causes of abortion. Complete records were sought of prior pregnancies, gynecologic surgery, non-steroidal estrogen exposure in utero and findings pathological examinations. The second group (control) composed of 100 women had at least one live birth with out pregnancy wastage.

**Setting :-** in the family planning in AL-Elwiya hospital and

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primary health center services in Bab-Almudhum.

Duration of study:- data collection continued for a period of one year starting on October 2011 and ending on October 2012.

Methods:- blood samples were collected using EDTA as anticoagulant to detect mycoplasma hominis and ureaplasma urealyticum using PCR technique.

DNA extraction:- blood samples were centrifuged at 3500 rpm for 20 min, cell pellets were treated with 20 mM Tris-HCl (pH 8.0), 1mM EDTA (pH 8.0), 5% SDS with 100 U $\mu$ g mL<sup>-1</sup> proteinase K. DNA was extracted using phenol-chloroform-isoamyl alcohol (25:24:1). The DNA was precipitated with 0.1 volume of 2 M potassium chloride and double volumes of absolute ethanol, air dried (8). The DNA pellets were suspended in TE buffer and incubated at 37°C for 30 min. Bioron \ Germany (Real line DNA extraction 2kit). PCR amplification:- Amplification of gene sequences was performed in a total volume of 50 mL of PCR buffer (10mM Tris-HCL, 50 mM KCL, pH 9) containing 0.1% Triton X-100, 200 mm each of dATP, dTTP, dGTP and dCTP, 100 pmol of each primer and 0.5 mg of DNA. purified mycoplasmal DNA(0.5-1 ng of DNA) was used as a positive control for amplification. The amplification was carried out for 40 cycles with denaturing at 94°C and annealing at 60°C Extension temperature was 72°C; finally, product extension was performed at 72°C for 10 min. Bioron \ Germany (Real line Flaformat kit).

Primers used as Mycoplasmal group specific (generic) PCR were prepared according to Ossewaarde et al. (9). as follows: upstream primer GPO-3,5- GGGG GCAA ACAG GATT AGAT TAGA TACC CT-3, downstream primer MGSO, 5-TGCAC CATC TGTC ACTC TGTT AACCTC-3 (expected size was 295 bp). Primers used in *Ureaplasma urealyticum* were prepared as follow : U.u. F1: 5- GCTAA TACCG AATAA TAACA TC-3 and U.u. R1:5-ATGGT ACAGT CAAAC TAAAA TC3 (expected size was 311 bp).

Statistical analysis:- Chi- square (X<sup>2</sup>) test was used for the generation of P<0.05 value.

### Result:

Results of PCR using primers specific for detection of mycoplasma are present in table (1). Out of 260 subjects, 87(33%) were positive for *M. homini* and/or *U. urealyticum* of these, 83 women were in the patient group (52%) and four were in the control group (2.5%). *M. homini* was isolated alone from 12\160 (7.5%) patients; no *M. homini* was detected in controls. The frequency of *U. urealyticum* detected alone in the patient group was 64\160(40%) and (4\100) controls (p<0.05). seven women within the patient group (4%) were co-infected with both *M. homini* and *U. urealyticum*.

**Table(1): Frequency of *Mycoplasma homini* and *Ureaplasma urealyticum* among patients and control womens.**

| Group            | Positive <i>Mycoplasma homini</i> (M.h) (%) | Positive <i>Ureaplasma urealyticum</i> (U.u) (%) | Positive mixed M.h and U.u (%) | Total (%) |
|------------------|---|--|--------------------------------|-----------|
| Patients (N=160) | 12 (7.5)                                    | 64 (40)  | 7 (4)                          | 83(52)    |
| Control (N=100)  | 0 (0)                                       | 4 (4)  | 0 (0)                          | 4(2.5)    |
| Total (N=260)    | 12 (5)                                      | 68 (26)  | 7 (3)                          | 87(33)    |

p<0.05

on comparing the frequency of genital mycoplasmas among the different age groups. We studied 160 female patients (pregnant loss) their ages ranged between the age of 15-49 years. The highest frequency of *M. homini* and *U. urealyticum* was seen in the age group of 20-24 years about 50% and 25% in age group 25-29 years for positive cases of *M. homini* and 36% were aged 25-29 years in positive cases of *U. urealyticum*, table (2). Significantly approximately two thirds of *M. homini* and *U. urealyticum* occurred in patients between the ages of 20 and 29 years (p<0.05).

**Table (2): Frequency of *Mycoplasma homini* (M.h) and *Ureaplasma urealyticum* (U.u) in different age groups.**

| Age range (years) | Total patient no. (%) | Positive cases |            |                       |
|-------------------|-----------------------|----------------|------------|-----------------------|
|                   |                       | M.h no.(%)     | U.u no.(%) | Mixed(M.h+U.u) no.(%) |
| 15-19             | 8 (5)                 | 0 (0)          | 1 (1.5)    | 1 (14)                |
| 20-24             | 32 (20)               | 6 (50)         | 10 (16)    | 4 (58)                |
| 25-29*            | 42 (26)               | 3 (25)         | 23 (36)    | 1 (14)                |
| 30-34             | 34 (21)               | 1 (8)          | 9 (14)     | 0 (0)                 |
| 35-39             | 17 (11)               | 2 (17)         | 12 (19)    | 1 (14)                |
| 40-44             | 21 (13)               | 0 (0)          | 6 (9)      | 0 (0)                 |
| 45-49             | 6 (4)                 | 0 (0)          | 3 (4.5)    | 0 (0)                 |
| Total (%)         | 160 (100)             | 12 (100)       | 64 (100)   | 7 (100)               |

\*p<0.05

Presence of genital mycoplasmas differ according to number of repeated abortions (table 3); such that *M. homini* was detected in the blood of cases suffering from three or four losses more than those with two or less pregnancy losses. Three cases out of 27 suffered from one abortion in addition to history of adverse outcome (neonatal death and/or intrauterine fetal death) were positive for *M. homini* (11%). Also *U. urealyticum* positive cases was detected from three repeated abortion (69%) more than two repeated abortions (19%). And 14 cases positive for *U. urealyticum* (52%) in cases suffered from one abortion in addition to history of adverse outcome. There was statistically significant difference between the frequency of *M. homini* and/or *U. urealyticum* and increase number of repeated abortion (p≤0.001).

**Table (3):** Frequency distribution of PCR-positive *Mycoplasma homini* and *Ureaplasma urealyticum* in relation to No. of pregnancy losses.

| PCR positive bacteria               | No. of pregnancy losses (160) |            |           |  | Total (%)    |
|-------------------------------------|-------------------------------|------------|-----------|--|--------------|
|                                     | No. of repeated abortion      |            |           | Causes suffered from abortion and adverse out come (%) * |              |
|                                     | ≥ 4 (%)                       | 3 (%)      | 2 (%)     |  |              |
| <i>Mycoplasma homini</i> (M.h)      | 3\56 (5)                      | 5\35 (14)  | 1\42 (2)  | 3\27 (11)  | 12\160 (7.5) |
| <i>Ureaplasma urealyticum</i> (U.u) | 18\56 (32)                    | 24\35 (69) | 8\42 (19) | 14\27 (52)   | 64\160 (40)  |
| Mixed M.h and U.u                   | 2\56 (4)                      | 3\35 (9)   | 1\42 (2)  | 1\27 (4)   | 7\160 (4)    |

p≤0.001

\* Adverse outcome still birth (intrauterine fetal death) and neonatal birth.

Spontaneous abortion at a gestation age of less than 36 weeks is shown (table 4). Of these 9(75%) cases had been colonized with *M. homini* and 50 (78%) with *U.urealyticum* in patient group. While in still birth (intrauterine fetal death) 3(25%) cases positive on *M. homini* and 11(17%) positive of *U. urealyticum*; also 3(5%) cases positive of *U. urealyticum* in neonatal death in patient group. Some of the patients were co-infected with *M. homini* and *U. urealyticum* (7 cases) as follows. Five cases with spontaneous abortion and 2 cases with intrauterine fetal death. The difference was statistically highly significant (p≤0.001).

**Table (4):** Number and percentages of pathological conditions observed in patients women and their possible association with mycoplasmas infection.

| Pathological conditions | Positive cases         |            |             |
|-------------------------|------------------------|------------|-------------|
|                         | Patient group (No.=76) |            |             |
|                         | * M.h (%)              | ** U.u (%) | M.h and U.u |
| Spontaneous abortion    | 9 (75)                 | 50 (78)    | 5 (71)      |
| Still birth             | 3 (25)                 | 11 (17)    | 2 (29)      |
| Neonatal death          | 0 (0)                  | 3 (5)      | 0 (0)       |
| Total (%)               | 12 (100)               | 64 (100)   | 7 (100)     |

p≤0.001

\*M.h= *Mycoplasma homini*

\*\* U.u= *Ureaplasma urealyticum*

**Discussion :**

The relation *mycoplasma hominis* and *Ureaplasma urealyticum* and repeated pregnancy loss was observed previously by several authors. In the present study, *M.hominis* was detected in blood of 12 (7.5%) women with repeated pregnancy losses, and not

in normal women (control), however (40%) of pregnancy loss women and (4%) in control group were colonized with *U.urealyticum*. 7 (4%) were infected with both *M.hominis* and *U.urealyticum*, where as none of the controls. As result detected by PCR technique. This is a little higher than results obtained by Elias et al. (10) who in a group of 222 women in similar age range, found *U.urealyticum* in (31.8 %) and *M.hominis* in only (3%) of the cases. Schlicht et al. (11) found a higher prevalence of 54% for *U.urealyticum*. Agbakoba (12) found a prevalence of (36.7%) for genital mycoplasmas while working with Nigerian women. Zdrodowska- stefanow et al. (13) in a similar study in pregnancy losses women, showed that *U.urealyticum* was detected in 161 (29.8%) and *M.hominis* in 20 (3.7%) women. Bayoumi et al. 2006 (1) concluded that the interaction between mycoplasmas and pro-inflammatory markers, which consist primarily of the development of specific antibody and non-specific interactions with B lymphocytes or antibody which responses are important in the resistance of mycoplasma disease in human. However, the ability of mycoplasmas to survive in their host despite vigorous responses suggests that these play a limited role in the host recovery from infection accounting for the appearance of systemic mycoplasma infection. In some cases, antibody response may contribute to the disease pathogenesis through the development to hypersensitivity responses or the deposition of immune complexes leading to auto immune reaction. Pandey et al 2005 (14) reported that majority of cases with unexplained cause of abortion are found to be associated with certain auto immune antibodies that may play a role in the immunologic failure in pregnancy and may lead to abortion. At present, the main method of detecting *M.hominis* and / or *U.urealyticum* is by culture, but the cultivation of mycoplasma infection is laborious, is time consuming, and requires specific expertise. PCR is revolutionizing the diagnosis of many infectious diseases, particularly those caused by organisms that

are difficult to cultivate. PCR is a more sensitive and reliable means of detecting *M.hominis* and *U.urealyticum* in blood and / or endocervical specimens; its results can be available within a day, compared with 2-5 days for culture. (6, 15) It was also observed that genital mycoplasma infection is specific to age group. The highest frequency was in the 20-29 years group with 9 (75%) with *M.hominis* and 33 (52%) in *U.urealyticum*. It is related to sexual activity, hence and sexual active group is a potential carrier. A similar result was obtained by Zdrodowska-Stefanow et al. (13) with the highest rate in the age range of 26-30 years (29.2% for *U.urealyticum* and 50.0% for *M.hominis*). The presence of genital mycoplasmas is associated with an increased risk of developing certain pathologic conditions of pregnancy, such as spontaneous abortion, preterm labor, and low birth weight. (2). In the present study, 75% of pregnant loss women infected with *M.hominis* and 78% infected with *U.urealyticum* in spontaneous abortion. More over 3/12 (25%) of *M.hominis* and 11/64 (17%) of *U.urealyticum* in still birth. While 3/64 (5%) of *U.urealyticum* in neonatal death. Cases had been colonized with *M.hominis* and /or *U.urealyticum*. This discrepancy may be due to variations in socioeconomic conditions and living standards. (2) since *M.hominis* and /or *U.urealyticum* has been found significantly associated with low socioeconomic background, such as poverty, number of sexual partners, and use of contraceptive drug. (16,17). A change in vaginal pH (e.g., bleeding in pregnancy, sexual intercourse, or vaginal douching) may predispose to an overgrowth of potential pathogens. (18) Although the precise role of *M.hominis* and /or *U.urealyticum* has been implicated in several complications of pregnancy and in neonatal morbidity and mortality. It may play roles in endometritis, chorioamnionitis, premature rupture of membranes, prematurity, low-birth weight infants, postpartum fever and it is an important cause of pneumonia and meningitis in very low-birth-weight infants. (6) Finally, we concluded, the prevalence of *M.hominis* and *U.urealyticum* infection was significantly correlated with the etiology of pregnancy losses. And we recommend that all women with poor pregnancy outcome, before planning subsequent pregnancy, should test for the presence of bacterial infections (Mycoplasma infection)

#### References:

1. Bayoumi, F.S.; Hussein, I.M.R and Hind, M.G., 2006. The role of mycoplasma infection and anticardiolipin antibodies as autoimmune parameters in pregnancy loss. *Journal of medical sciences*, 6: 585-590.
2. Bayraktar, M.R.; Ozerol, I.H.; Gucluer, N.; Celik, O., 2009. Prevalence and antibiotic susceptibility of *Mycoplasma hominis* and *Ureaplasma urealyticum* in pregnant women. *Journal of medical microbiology*, 3:10-20
3. Daxboeck, F.; Zitta, S.; Stadler, M.; Iro, E., Karause, R. 2005. *Mycoplasma hominis* and *Ureaplasma urealyticum* in patients with sterile pyuria. *Journal of infectious*, 51:54-58.
4. Patai, K.; Szilagy, G.; Szentmariay, I.F; Paulin, F., 2005. Severe endometritis caused by genital mycoplasmas after cesarean section. *Journal of medical microbiology*, 54:1249-1250.
5. Witt, A.; Berger, A.; Gruber, C.J.; Petricevic, L.; Apfalter, P.; Worda, C., 2005. Increased intrauterine frequency of *Ureaplasma urealyticum* in women with preterm labor and preterm premature rupture of the membranes and subsequent cesarean delivery. *Am. J. Obstet. Gynecol*, 193:1663-1669.
6. Peerayeh, S.N and Samimi, R.Y., 2007. Detection of *Ureaplasma urealyticum* in clinical samples from infertile women by polymerase chain reaction. *Iranian journal of pharmacology and therapeutics*. 6.1: 23-26.
7. Kuppeveld, F.J. M.; Logt, J. T. M.; Angulo, A. F., 1992. Genus and species-specific identification of mycoplasmas by 16SrRNA amplification. *Appl. Env. Microbiol*. 52: 2606-2615.
8. Ghadersohi, A. R. J.; Coelen, A. and Hirst, R. G. F. 1997. Development of specific DNA probe and PCR for the detection of *Mycoplasma bovis*. *Vet. Microbiol. J*. 56:87-98.
9. Ossewaarde, J. M., Devries, A., Bestebroer, T. and Angulo A. F., 1996. Application of mycoplasma group-specific PCR for monitoring decontamination of mycoplasma infected *Chlamydia Sp. Strains*. *Applied. Environ. Microbiol*. 62:328-331.
10. Elias, M.; Grazesko, J.; Siejkowski, R., 2005. Obecność *Mycoplasma hominis* and *Ureaplasma urealyticum* w kanale szyjki macicy kobiet. *Gin. Pol*. 76:28-32.
11. Schlicht, M. J.; Lovrich, S. D.; Sartin, J. S. 2004. High prevalence of genital mycoplasmas among sexually active young adults with urethritis or cervicitis symptoms in Lacrosse, Wisconsin. *J. Clin. Microbiol*. 42:4636-4640.
12. Agbakoba, N. R., 2007. Prevalence of mycoplasma and ureaplasma in women attending gynaecology clinic at university college hospital, Ibadan and pathogenicity of *Ureaplasma urealyticum* in mice. Ph. D. thesis, University of Ibadan Nigeria.
13. Zdrodowska-Stefanow, B.; Klosowska, W. M., Ostaszewska-puchalska, I., 2006. *Ureaplasma urealyticum* and *Mycoplasma hominis* infection in women with urogenital diseases. *Adv. Med. Sci*, 51: 250-254.
14. Pandey, M.C.; Rani, R. and Agrawal, S., 2005. An update in recent spontaneous abortion. *Arch. Gynecol. Obstet*. 272:95-108.
15. Leon, X.; Blanchard, A.; and Henstehel, A., 1994. Comparison of PCR with culture for detection of *Ureaplasma urealyticum* in clinical samples from patients with urogenital infections. *J. Clin. Microbiol*. 32:2232-2234.
16. Domingues, D.; Tavira, L. T.; Durate, A. 2003. Genital

mycoplasmas in women attending a family planning clinic in Gwine-Bissau and their susceptibility to antimicrobial agents. Acta. Tropica. 86:19-24.

17. Keane, F. E; Thoma, B. J.; Gilroy, C. B. 2000. The association of Mycoplasma homini , ureaplasma urealyticum, and mycoplasma genitalium with bacterial vaginosis: bservation

on hetosexual women and their male partners. Int. J. STD. AIDS. 11: 356-360.

18. Pararas, M.V; Skevaki, C.L., kafetzis, D.A. 2006. Preterm birth due to maternal infection: causative pathogens and modes of prevention. Eur. J. Clin. Microbiol. Infect. Dis. 25:562-569.