

Long-Term Effects of Scopolamine on Brain Tissue of Mice

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Abstract:

Background: Scopolamine is an anticholinergic drug that disrupts cholinergic transmission in the central nervous system as well as causes cognitive abnormalities and pathological hallmarks that are similar to those seen in Alzheimer's Disease. Therefore, it is used for induction of Alzheimer's Disease in animal models.

Objective: to investigate the effects of long-term induction with scopolamine on the brain tissue of mice.

Methods: Seventy adult mice were divided into 2 equal groups: The first group was the normal control group received distilled water only. The second one was the Alzheimer's Disease induction group received intraperitoneal scopolamine (1mg/kg) for 14 days only after that distilled water was given for the next 6 months. Ten mice were isolated from each group at zero time, after 2 weeks of induction, after 3-month and after 6 months and subjected to the behavioral tests then sacrificed for determination of biochemical factors (including brain-derived neurotrophic factor, total antioxidant status, malondialdehyde, and amyloid β). Data were analyzed using *t*-tests, and ANOVA. All values expressed as Mean \pm SD and *P* value <0.05 were considered significant.

Result: Scopolamine produced brain histopathological changes similar to those of human Alzheimer's disease. However, it does not produce further statistically significant differences in behavioral tests and biochemical markers during the total period of study.

Conclusion: scopolamine produces brain tissue changes that persist for a long period and it can be used for long-term study of Alzheimer's disease.

Keywords: Alzheimer's disease; Antioxidant; Cognitive function; Oxidative stress; Scopolamine.

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Introduction

Scopolamine (SCM) is an anticholinergic drug that disrupts cholinergic transmission in the central nervous system (CNS) and demonstrates competitive antagonism at muscarinic acetylcholine receptors (mAChRs) (1).

As it easily crosses the blood-brain barrier, scopolamine is frequently utilized in neuroscience research to induce cognitive impairments, similar to those seen in Alzheimer's Disease (AD), in experimental animals (2). These animal models are commonly used to test medications for possible therapeutic usefulness in people with AD-type dementia (1, 3).

Moreover, it is used with different doses in different studies. Some of these studies used it as a single dose and others as multiple doses. Therefore, it is unclear if SCM can induce different intensities and durations of amnesia after single or repeated doses (4).

In addition, many studies indicated that SCM increases the deposition of $A\beta$, the level of reactive oxygen species, and reduces antioxidant concentration leading to lipid peroxidation that induces oxidative stress.

Also, it decreases the brain-derived neurotrophic factor (BDNF) and cAMP-response element binding protein (CREB) expression in the brain which are hallmarks of AD disease and ultimately cause memory impairment and synaptic dysfunction (3, 5). Previous studies concerned with the use of SCM for induction of AD in animal models did that over a short induction period. Therefore, the current study aimed to investigate the long-term effects of scopolamine on cognitive and memory functions as well as the pathological hallmark of Alzheimer's disease in mice.

Methods

An experimental study was conducted from 1st, July 2022 to 1st, June 2023.

The current study involved ninety adult female mice (4–8 weeks old) weighing (20–25g) were purchased from the Al-Razi center, Ministry of Industry and Minerals, Baghdad, Iraq. The mice were kept in the experimental area for 2 weeks for the habituation phase and housed as ten mice per cage at an appropriate temperature (25°C) and humidity (30% \pm 10%), with a standard 12-hour light/dark cycle and free access to water and standard food (high protein feed and milk powder).

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In the beginning, twenty mice were included in a pilot study to detect the effective dose and duration of treatment for Scopolamine Intraperitoneal (IP/SCM) to induce AD (scopolamine hydrochloride, 1mg/kg, was dissolved in distilled water for injection and given IP to induce AD in mice (6). The results of the pilot study showed that IP/SCM (1mg/kg) for 14 days induced AD effectively. Then, seventy adult mice were divided into 2 equal groups; the first group was the normal group that received distilled water only during the total period of the study and was considered a control group, and the second one was the induction untreated Alzheimer group that received IP/SCM (1 mg/kg) for 14 days only after that distilled water was given for the next 6 months.

Mice were subjected to behavioral tests (Barnes Maze (7), Novel Object Recognition (8), and Y-Maze Tests (9). Then, these mice were anesthetized and sacrificed to isolate brain tissue for further determination of biochemical markers such as BDNF, TAS, MDA, and 1-42 β -amyloid peptide. This procedure was repeated 3 and 6 months after induction.

Data were analyzed using *t*-tests (paired and unpaired) and ANOVA. All values expressed as Mean \pm SD and *P* value <0.05 were considered significant (10).

Results

Histopathological changes

Results from the current study revealed that histopathological sections of brain tissue from mice induced with SCM for 2 weeks showed microscopically changes (Figures 1 & 2)

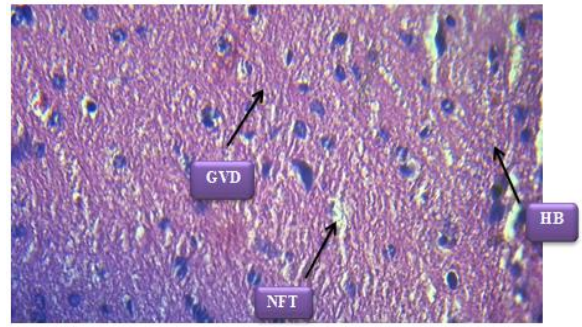


Figure (1): Histological section of mice AD brain tissue after induction with scopolamine for 2 weeks (current study) showing numerous NFT, HB, and GVD (H&E stain) (40X).

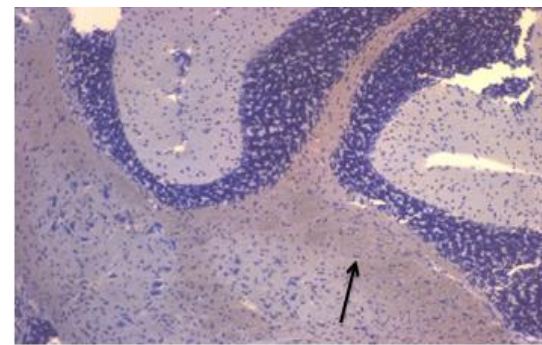


Figure (2): Histological section of mice AD brain tissue after induction with scopolamine for 2 weeks (current study) showing multifocal severe deposition of amyloid beta plaques (orange-red color) (arrow) (Congo red stain) (10X).

Also, findings indicated the validity of AD animal model created in the current study. Similar to those reported in human AD brain tissue (Figures 3, 4, and 5)

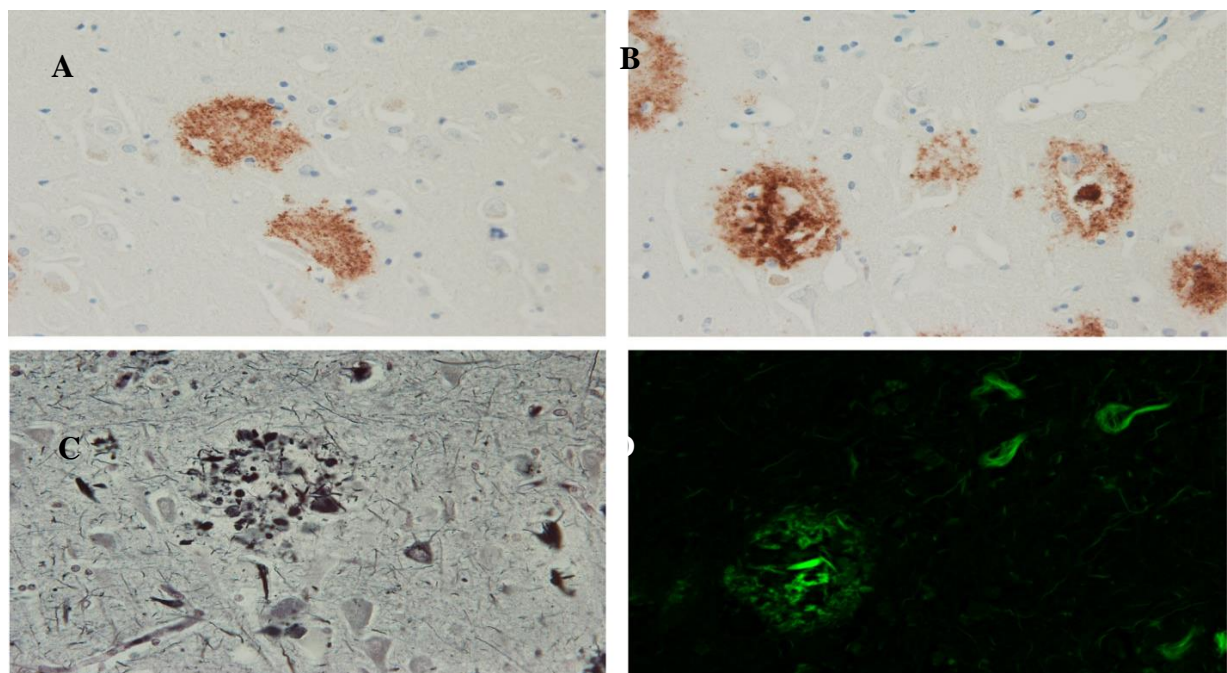


Figure (3): Histological section of human AD brain tissue showing the presence of amyloid β Senile Plaques by using antibodies directed against A β peptides (A&B), Bielschowsky silver staining (C) or Thioflavin S staining (D) (DeTure & Dickson, 2019).

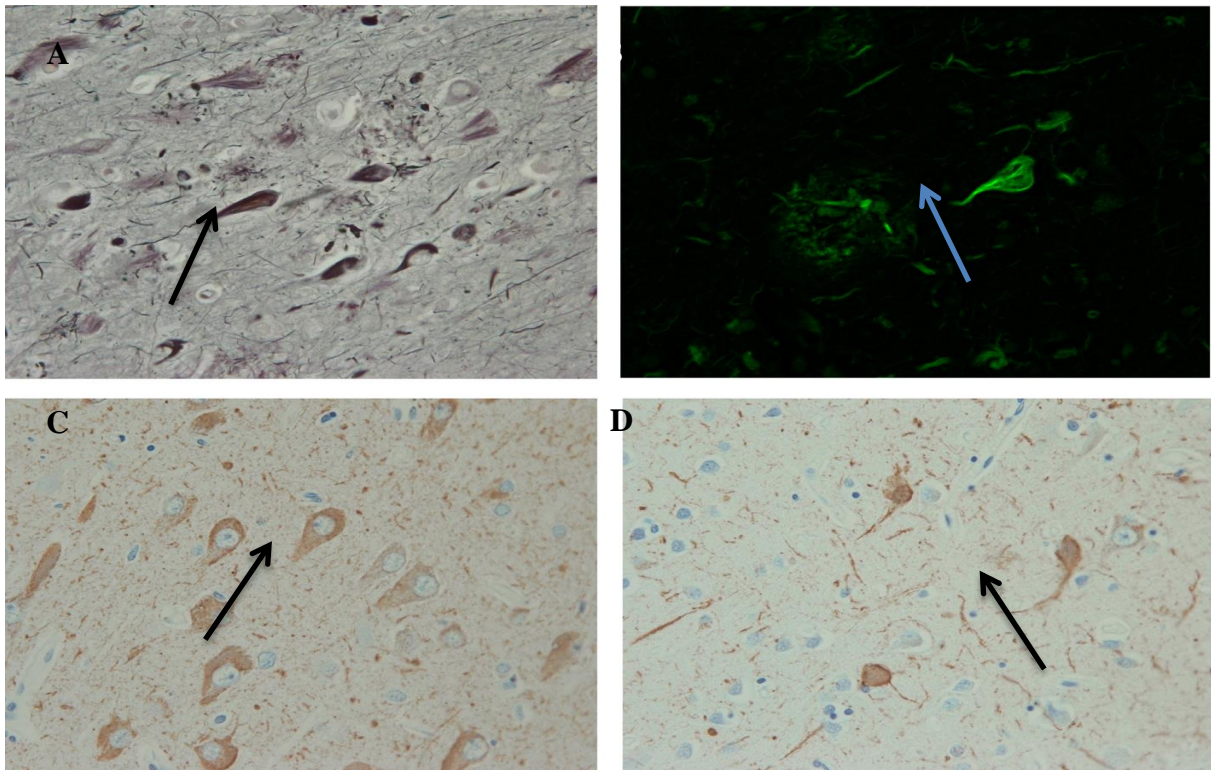


Figure (4): Histological section of human AD brain tissue showing the presence of Neurofibrillary Tangles by using Silverstaining (A), Thioflavin S (B), and tau immunohistochemistry (C, D). (DeTure & Dickson, 2019).

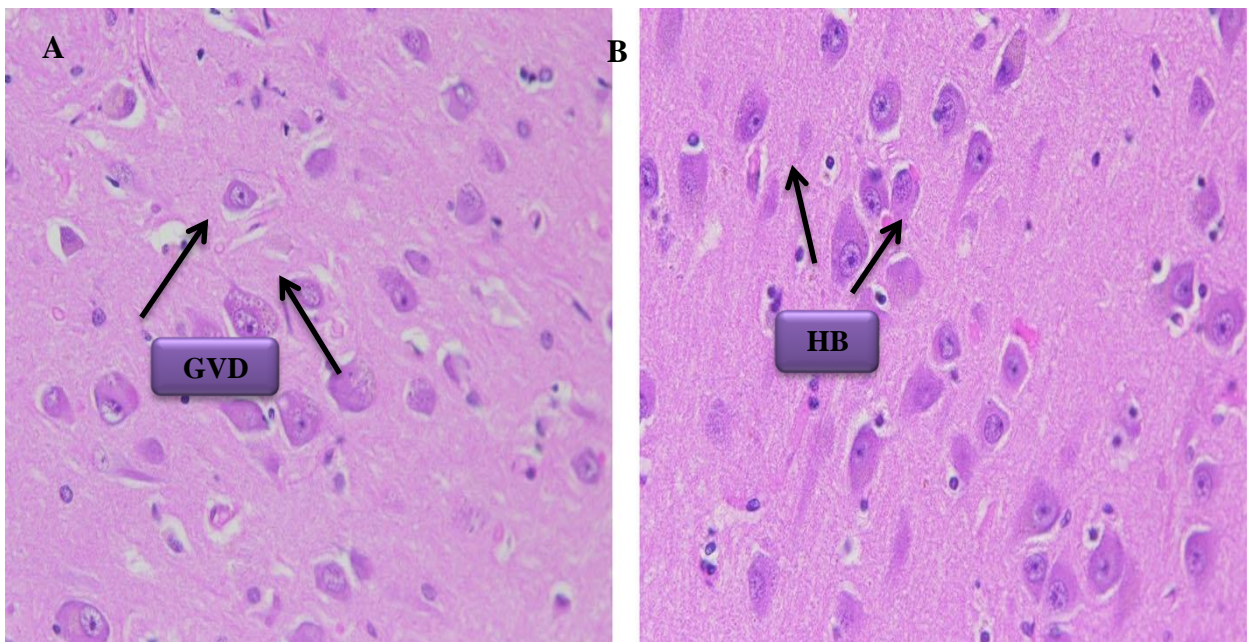


Figure (5): Histological section of human AD brain tissue showing the presence of Granulovacuolar Degeneration (acid phosphatase histochemistry) (A) and Hirano Bodies (H&E stain) (B) (DeTure & Dickson, 2019).

Behavioral tests and biochemical markers along the total period of the study for normal control groups: The current study revealed that there were no statistically significant differences in behavioral tests (Barnes Maze Test, Novel Object Recognition Test, and Y-Maze Test) or biochemical markers (Amyloid β , BDNF, MDA, and TAS) in the normal control groups throughout the entire duration of study (6 months) (Tables 1 & 2, respectively).

Table (1): Behavioral tests for normal groups at zero time, after 3 months, and after 6 months of treatment (Y-maze Test, Novel Object Recognition Test, and Barnes Maze Test)

| Behavioral Test | Mean ± SD | | | P value | | |
|------------------|----------------------|-----------------------|----------------|---------------|-------------|-------|
| | Nor. 0T (n=10) | Nor. 3M (n=10) | Nor. 6M (n=10) | | | |
| Y- Maze Test | 79.346±3.774 | 77.554±4.816 | 77.384±2.215 | 0.681 | | |
| NOR Test | 80.70±4.446 | 76.506±2.073 | 78.153±6.057 | 0.341 | | |
| Barnes Maze test | Acquisition training | Primary latency(s) | 45.7±6.056 | 41.5±4.428 | 42±5.657 | 0.196 |
| | | Primary error | 16±3.367 | 17±4.522 | 20±5.888 | 0.210 |
| | | Primary hole distance | 8.2±2.699 | 9.3±2.750 | 8.875±1.458 | 0.603 |
| | Probe trial | Primary latency(s) | 15.8±3.564 | 16.6±2.881 | 16.2±3.899 | 0.936 |
| | | Primary error | 6±1.581 | 5.8±1.924 | 4.4±1.342 | 0.278 |
| | | Primary hole distance | 4.4±1.140 | 6.2±1.789 | 8±2.916 | 0.055 |
| | Reversal learning | Primary latency(s) | 50.555±7.859 | 54.142±11.889 | 51.4±12.176 | 0.715 |
| | | Primary error | 13.111±3.919 | 10.8±2.932 | 12±2.748 | 0.231 |
| | | Primary hole distance | 10.555±2.455 | 11.062±2.909 | 10.2±2.529 | 0.722 |

Nor. 0T=normal group at zero time, Nor. 3M = normal group after 3 months of treatment, Nor. 6M = normal group after 6 months of treatment, s = second, SD = Standard deviation, NOR = Novel Object Recognition Test.

Tables (2): Mean levels of biochemical markers for normal control groups at zero time, normal after 3 months, and normal after 6 months of treatment

| Marker | Mean ± SD | | | P- value |
|------------------|----------------|----------------|-----------------|----------|
| | Nor. 0 T (n=6) | Nor. 3 M (n=6) | Nor. 6 M (n=6) | |
| BDNF (ng/mL) | 3.062 ± 0.966 | 4.092 ± 1.134 | 4.026 ± 1.125 | 0.354 |
| Amyloid β (ug/L) | 9.930 ± 0.567 | 12.207 ± 1.683 | 11.694 ± 1.846 | 0.076 |
| TAS (pg/mL) | 35.192± 3.672 | 37.378 ± 1.02 | 41.887 ± 10.994 | 0.096 |
| MDA (nmol/mL) | 0.621 ± 0.018 | 0.613 ± 0.05 | 0.594 ± 0.117 | 0.845 |

Nor. 0T=normal group at zero time, Nor. 3M = normal group after 3 months of treatment, Nor. 6M = normal group after 6 months of treatment, BDNF= Brain derived neurotrophic factor, TAS = Total antioxidant status, MDA = Malondialdehyde, SD = Standard deviation.

Behavioral tests and biochemical markers for AD induction groups over the total period of the study: Current study revealed that there were no statistically significant differences in behavioral tests (Barnes Maze Test, Novel Object Recognition Test,

and Y-Maze Test) or biochemical markers (Amyloid β, BDNF, MDA, and TAS) during the total period of the study for the AD-induction groups (using SCM) after 2 weeks, 3 months, and 6 months (Tables 3 and 4, respectively).

Table (3): Behavioral tests for AD induction groups after 2 weeks, 3 months, and 6 months of treatment

| Behavioral Test | Mean ± SD | | | P- value | |
|----------------------|---------------------------|---------------------------|---------------------------|----------------|-------|
| | Induction (SCM 2W) (n=10) | Induction (SCM 3M) (n=10) | Induction (SCM 6M) (n=10) | | |
| Y- Maze Test | 46.44±5.185 | 48.424±3.704 | 51.204±2.224 | 0.751 | |
| NOR Test | 45.744±2.196 | 47.268±1.846 | 46.31±2.957 | 0.605 | |
| Acquisition training | Primary latency (s) | 158.667±26.585 | 157.25±21.57 | 148.111±29.118 | 0.598 |
| | Primary error | 37.833±9.637 | 38±8.569 | 36.555±5.659 | 0.912 |
| | Primary hole distance | 17.166±1.642 | 17.733±1.869 | 18±2.397 | 0.597 |
| Probe trial | Primary latency (s) | 58.5±1.914 | 57.5±3.507 | 57±5.196 | 0.846 |
| | Primary error | 19±3 | 22.833±3.544 | 20.2±2.280 | 0.137 |
| | Primary hole distance | 16.2±2.863 | 17.667±2.250 | 15.6±2.701 | 0.416 |
| Reversal learning | Primary latency (s) | 143.625±17.508 | 146.8±25.952 | 165.75±16.568 | 0.092 |
| | Primary error | 34±7.406 | 31±6.96 | 30.75±7.648 | 0.611 |
| | Primary hole distance | 16.5±3.070 | 17.4±2.633 | 17.75±2.187 | 0.627 |

SCM 2W = after 2 weeks of induction by scopolamine, SCM 3M= after 3 months of treatment in the scopolamine induction group, SCM 6M= after 6 months of treatment in the scopolamine induction group, s = second, SD = Standard deviation, NOR = Novel Object Recognition Test.

Table (4): A comparison of the mean levels of biochemical markers in the induction groups throughout the study

| Marker | Mean±SD | | | P value |
|------------------|--------------------------|--------------------------|--------------------------|---------|
| | Induction (SCM 2W) (n=6) | Induction (SCM 3M) (n=6) | Induction (SCM 6M) (n=6) | |
| BDNF (ng/mL) | 0.686 ± 0.137 | 0.624 ± 0.213 | 0.643 ± 0.161 | 0.832 |
| Amyloid β (ug/L) | 57.227± 8.553 | 61.827± 10.059 | 49.122± 11.292 | 0.062 |
| TAS (pg/mL) | 10.123 ± 2.949 | 11.336 ± 1.042 | 9.450 ± 3.092 | 0.438 |
| MDA (nmol/mL) | 2.764 ± 0.344 | 2.560 ± 0.377 | 3.012 ± 0.763 | 0.354 |

Table (5): Comparison between the AD induction group and normal control group at zero time for behavioral (Y-maze, Novel Object Recognition, and Barnes Maze) tests

| Behavioral Test | Mean±SD | | P-value | | | |
|------------------|----------------------|---------------------------|-----------------------|----------------|------------|--------|
| | Nor. 0T (n=10) | Induction (SCM 2W) (n=10) | | | | |
| Y maze test | 79.346±3.773 | 46.44±5.185 | <0.001 | | | |
| NOR Test | 80.70±4.446 | 45.744±2.196 | <0.001 | | | |
| Barnes Maze test | Acquisition training | Primary latency (s) | 45.7±6.056 | 158.666±26.585 | <0.001 | |
| | | Primary error | 16±3.366 | 37.833±9.637 | <0.001 | |
| | | Primary hole distance | 8.2±2.699 | 17.166±1.642 | <0.001 | |
| | | Probe trial | Primary latency (s) | 15.8±3.563 | 58.5±1.914 | <0.001 |
| | | | Primary error | 6±1.581 | 19±3 | <0.001 |
| | | | Primary hole distance | 4.4±1.14 | 16.2±2.863 | <0.001 |
| | Reversal learning | Primary latency (s) | 50.555±7.859 | 143.625±17.508 | <0.001 | |
| | | Primary error | 13.111±3.919 | 34±7.406 | <0.001 | |
| | | Primary hole distance | 10.555±2.455 | 16.5±3.07 | <0.001 | |

SCM 2W = after 2 weeks of induction by scopolamine, Nor. 0T=normal group at zero time, s = second, SD = Standard deviation, NOR = Novel Object Recognition Test.

Comparison of Biochemical markers between the AD induction group after 2 weeks and the Normal control group at zero time: The current study indicated that there was a highly significant elevation in the mean level of amyloid β and MDA, while there was a significant reduction in the mean level of BDNF and TAS in the AD induction group after 2 weeks in comparison with the normal control group at zero time (Table 6).

Table (6): Comparison of mean levels of biochemical markers between AD induction groups and normal group at zero time

| Marker | Mean±SD | | P-value |
|------------------|----------------|--------------------------|---------|
| | Nor. 0 T (n=6) | Induction (SCM 2W) (n=6) | |
| BDNF(ng/mL) | 3.062 ± 0.966 | 0.686 ± 0.137 | 0.006 |
| Amyloid β (ug/L) | 9.930 ± 0.567 | 57.227± 8.553 | <0.001 |
| TAS (pg/mL) | 35.192± 3.672 | 10.123 ± 2.949 | <0.001 |
| MDA (nmol/ml) | 0.621 ± 0.018 | 2.764 ± 0.344 | 0.003 |

SCM 2W = after 2 weeks of induction by scopolamine, Nor. 0T=normal group at zero time, SD = Standard deviation, BDNF= Brain derived neurotrophic factor, TAS = Total antioxidant status, MDA = Malondialdehyde.

Discussion

Validity of the animal model of AD created in the current study: In the current study, SCM caused accumulation of amyloid (Aβ) plaques, NFT, GVD,

and HB by histopathological examination, which is similar to AD of human brain tissue (11). The similarities between human and mouse AD brain tissues obtained in the current study support the validity of the animal model created in the current study and, therefore, suggest that its results can be applied to humans with AD (12). Behavioral tests and biochemical markers for normal control groups: The age of the young mice enrolled in the current study at zero time was (1-2 months), while after 3 months of therapy was (4-5 months), and after 6 months of therapy was (7-8 months). In this study, the cognitive and memory functions of normal control mice were preserved over the total period of the study by measuring behavioral tests including the Barnes maze test, Novel Object Recognition (NOR) Test, and Y-maze test. Thus, these data excluded any effect of aging on the result of the current study (13). These results were in agreement with the results of some previous studies that assessed age-related memory and cognitive function in multiple age groups of normal mice (14-17). Aging is associated with cognitive decline and may be linked to minimal neuronal loss (13). However, mature CNS neurons at a young age are very resistant to cell death. Therefore, neuronal cell death is limited to the adult brain (18). Additionally, the present study observed that there were no differences in the level of Aβ in the brains of normal mice over the total period of the study and these results agreed with those from previous studies

Comparison of behavioral tests between the AD-induction group after 2 weeks and the normal control group at zero time

that measured the level of $A\beta$ in the brains of normal mice (19, 20). Furthermore, for different ages of mice (young, middle-aged, and elderly), the level of the β -secretase (BACE1) enzyme that is necessary for the synthesis of $A\beta$ is consistent (19). Consequently, the $A\beta$ level during the young age is consistent. The present study found that there are no differences in the levels of BDNF in young normal mice of different ages throughout the study. These findings agreed and contrasted those of some previous studies (21-24). The BDNF gene plays a crucial role in neuronal generation, function, and memory. In addition, neuronal BDNF mRNA expression of the hippocampus and cortex is unaffected by aging in normal mice, hence the level of BDNF remains consistent over time (25). Regarding oxidative stress level, the current study has shown that there have been no statistically significant changes in the concentration of MDA and TAS among young normal mice of varied ages. One of the earliest studies has compared the activity of antioxidant enzymes and MDA levels among normal mice of different ages and discovered a contradicter result in which GPx and copper-zinc superoxide dismutase CuZn/SOD activities are higher in 18 and 28 months old mice than younger, 2 month old, mice, while manganese superoxide dismutase (MnSOD), GRD activities, and MDA level do not change with aging (26).

Oxidative damage is considered one of the predominant mechanisms of cellular and tissue damage in aging (27). The oxidative damage during aging occurs due to neuroinflammation, and increased expression of pro-inflammatory factors (28). In the healthy brain of young mice, microglia are "resting" and dormant, which eliminates the impact of neuroinflammation on young mice's normal brain function (29). Thus, OS in young mice is kept at a low level.

Biochemical markers and behavioral tests for AD-induction group over the total period of the study: The current study revealed that the effects of SCM on cognitive and memory function as well as biochemical markers (Amyloid β , BDNF, MDA, and TAS) were consistent throughout the study (6 months). These results suggested that SCM has a prolonged effect, which will strengthen the impact of the study's medications and these findings have not been reported in other previous studies because all other studies focused on the short-term effects of SCM.

Scopolamine causes cholinergic neuronal damage in the hippocampus by enhancing DNA damage and inhibiting the mRNA expression of many genes encoding neuronal factors that are crucial for cell survival as well as increasing oxidative stress by enhancing lipid peroxidation and decreasing the antioxidant system capacity (30). Additionally, SCM

increases $A\beta$ deposition which further enhances oxidative stress (31). Furthermore, SCM interferes with the expression of neurofilaments, which are essential for axonal transport in neurons (32). The loss of cholinergic function in the hippocampus is associated with serious cognitive impairments (32) which may last for a long time.

On the other hand, results obtained from this study found that SCM interfered with, and subsequently caused impairment of, learning and short-term as well as long-term memories as evidenced by conducting Barnes-maze, NOR, and Y-maze tests.

It has been demonstrated that SCM can impair memory and cognitive functions in mice by administering single or multiple injections at various concentrations through assessing many behavioral tests (33-36).

Aykac et al. (2018) found that administering SCM (1mg/kg) IP for 14 days dramatically raised MDA levels, lowered GSH levels, decreased BDNF expression, and lowered short- and long-term memory (36). In addition, Anand et al. (2022) concluded that SCM single injection (2mg/kg) increased oxidative stress in the hippocampus by increasing thiobarbituric acid-reactive substances (TBARS), which reflect lipid peroxidation and decreased GSH as well as CAT levels (37). Also, a recent study conducted by Cheedella et al. (2023) found that SCM (5mg/kg) for 7 days reduced CAT activity and H&E-stained histological sections of the brain showed severe blood capillary congestion with perivascular edema (scars), along with edema and deposition of amyloid plaques in the hippocampus when compared with normal mice (31).

However, a study done by Ban et al. (2020) found conflicting results regarding the impact of SCM on oxidative stress such as that CAT activity was unaffected after SCM injection in mice compared to normal mice (34). Additionally, Lee et al. (2010) found that SCM significantly increased SOD and GSH-Px antioxidant enzyme activities (38).

Despite that, it is well-known that the non-selective muscarinic acetylcholine receptor antagonist scopolamine (SCM) prevents cholinergic signals from traveling through the brain. This SCM-induced dysregulation of cholinergic activity and increased activity of acetylcholinesterase in the hippocampus interferes with mouse learning and memory functions (3, 32). One of the mechanisms causing scopolamine-induced amnesia is oxidative stress through increasing the levels of malondialdehyde, a marker of lipid peroxidation, and lowers the activity of many antioxidant enzymes (4). Additionally, there is a relationship between $A\beta$ and oxidative stress, because prooxidants elevate $A\beta$ formation and $A\beta$ generates oxidative stress (39). Moreover, the degree of synapse loss and cognitive deficits do correlate well with the amounts of soluble $A\beta$ in the brain (19).

AD brain tissue. In addition, the effects of scopolamine on memory and cognitive functions as well as on pathological hallmark of AD persist for a long period (about 6 months).

Conclusion

In light of results reported by the current study, scopolamine produces histopathological changes in mice brain tissue similar to those reported in human

Authors' Declarations

Mohammed AH AL- Zobaidy is an Editorial board member but did not participate in the peer review process other than as an author.

We hereby confirm that all the Figures and Tables in the manuscript are ours. The project was approved by the local ethical committee at the Department of Pharmacology/ College of Medicine, University of Baghdad, Baghdad, Iraq, with reference number (PHARMACOMED) in U.vB 23.13 (Appendix I).

Conflict of interest: None

Funding Source: None

Authors' contribution

Study conception & design Mohammed AH JABarah AL- Zobaidy. Literature search & Manuscript preparation are done by Neven Nihal Hana Istifo. Data acquisition, Data analysis & interpretation are done by Neven Nihal Hana Istifo and Kasim Sakran Abass. Manuscript editing & review are done by Mohammed AH JABarah AL- Zobaidy.

References

- Cheon SY, Koo B-N, Kim SY, Kam EH, Nam J, Kim EJ. Scopolamine promotes neuroinflammation and delirium-like neuropsychiatric disorder in mice. *Scientific Reports*. 2021;11(1):8376. <https://doi.org/10.1038/s41598-021-87790-y>.
- Chen WN, Yeong KY. Scopolamine, a toxin-induced experimental model, is used for research in Alzheimer's disease. *CNS & Neurological Disorders-Drug Targets (Formerly Current Drug Targets-CNS & Neurological Disorders)*. 2020;19(2):85-93. <https://doi.org/10.2174/1871527319666200214104331>.
- Yadang FSA, Nguzeze Y, Kom CW, Betote PHD, Mamat A, Tchokouaha LRY, et al. Scopolamine-induced memory impairment in mice: neuroprotective effects of *Carissa edulis* (Forssk.) Valh (Apocynaceae) aqueous extract. *International Journal of Alzheimer's Disease*. 2020;2020. <https://doi.org/10.1155/2020/6372059>.
- Rahimzadegan M, Soodi M. Comparison of memory impairment and oxidative stress following single or repeated doses administration of scopolamine in rat hippocampus. *Basic and clinical neuroscience*. 2018;9(1):5. <https://doi.org/10.29252/NIRP.BCN.9.1.5>.
- Hernández-Rodríguez M, Arciniega-Martínez IM, García-Marín ID, Correa-Basurto J, Rosales-Hernández MC. Chronic administration of scopolamine increased GSK3 β P9, beta secretase, amyloid beta, and oxidative stress in the hippocampus of Wistar rats. *Molecular Neurobiology*. 2020;57:3979-88. <https://doi.org/10.1007/s12035-020-02009-x>.
- Mahdi O, Baharuldin MTH, Nor NHM, Chiroma SM, Jagadeesan S, Moklas MAM. Chemicals used for the induction of Alzheimer's disease-like cognitive dysfunctions in rodents. *Biomedical Research and Therapy*. 2019;6(11):3460-84. <https://doi.org/10.15419/bmrat.v6i11.575>.
- Pitts MW. Barnes maze procedure for spatial learning and memory in mice. *Bio-protocol*. 2018;8(5):e2744-e. <https://doi.org/10.21769/BioProtoc.2744>.
- Lueptow LM. Novel object recognition test for the investigation of learning and memory in mice. *JoVE (Journal of Visualized Experiments)*. 2017(126):e55718. <https://doi.org/10.3791/55718>.
- Prieur EA, Jadavji NM. Assessing spatial working memory using the spontaneous alternation Y-maze test in aged male mice. *Bio-protocol*. 2019; 9(3): e3162-ed <https://doi.org/10.21769/BioProtoc.3162>.
- Kirkwood BR, Sterne JA. *Essential medical statistics*: John Wiley & Sons; 2010.
- DeTure MA, Dickson DW. The neuropathological diagnosis of Alzheimer's disease. *Molecular neurodegeneration*. 2019;14(1):1-18. <https://doi.org/10.1186/s13024-019-0333-5>.
- Miller JA, Horvath S, Geschwind DH. Divergence of human and mouse brain transcriptome highlights Alzheimer disease pathways. *Proceedings of the National Academy of Sciences*. 2010;107(28):12698-703. <https://doi.org/10.1073/pnas.0914257107>.
- Radulescu CI, Cerar V, Haslehurst P, Kopanitsa M, Barnes SJ. The aging mouse brain: cognition, connectivity and calcium. *Cell Calcium*. 2021;94:102358. <https://doi.org/10.1016/j.ceca.2021.102358>.
- Shoji H, Miyakawa T. Age-related behavioral changes from young to old age in male mice of a C57 BL/6J strain maintained under a genetic stability program. *Neuropsychopharmacology reports*. 2019;39(2):100-18. <https://doi.org/10.1002/npr2.12052>.
- Hendrickx JO, De Moudt S, Calus E, De Deyn PP, Van Dam D, De Meyer GR. Age-related cognitive decline in spatial learning and memory of C57BL/6J mice. *Behavioural brain research*. 2022; 418: 113649. <https://doi.org/10.1016/j.bbr.2021.113649>.
- Crespo NE. Age-Related Cognitive Decline in Female C57BL/6cnp Mice 15-16 Months of Age. *J Biomed Eng*. 2021; 5: 1-12. <https://doi.org/10.17303/jber.2021.5.101>.
- Clifford KP, Miles AE, Prevot TD, Misquitta KA, Ellegood J, Lerch JP, et al. Brain structure and working memory adaptations associated with maturation and aging in mice. *Frontiers in Aging Neuroscience*. 2023; 15. <https://doi.org/10.3389/fnagi.2023.1195748>.
- Chi H, Chang H-Y, Sang T-K. Neuronal cell death mechanisms in major neurodegenerative diseases. *International journal of molecular sciences*. 2018; 19 (10): 3082. <https://doi.org/10.3390/ijms19103082>.
- Janssen L, Keppens C, De Deyn PP, Van Dam D. Late age increase in soluble amyloid-beta levels in the APP23 mouse model despite steady-state levels of amyloid-beta-producing proteins. *Biochimica et Biophysica Acta (BBA)-Molecular*

- Basis of Disease. 2016;1862(1):105-12. <https://doi.org/10.1016/j.bbadis.2015.10.027>.
20. Ameen-Ali KE, Simpson JE, Wharton SB, Heath PR, Sharp PS, Brezzo G, et al. The time course of recognition memory impairment and glial pathology in the hAPP-J20 mouse model of Alzheimer's disease. *Journal of Alzheimer's Disease*. 2019;68(2):609-24. <https://doi.org/10.3233/JAD181238>.
21. Endres T, Lessmann V. Age-dependent deficits in fear learning in heterozygous BDNF knock-out mice. *Learning & memory*. 2012;19(12):561-70. <http://www.learnmem.org/cgi/doi/10.1101/lm.028068.112>.
22. Psotta L, Lessmann V, Endres T. Impaired fear extinction learning in adult heterozygous BDNF knock-out mice. *Neurobiology of learning and memory*. 2013;103:34-8. <https://doi.org/10.1016/j.nlm.2013.03.003>.
23. Harb M, Jagusch J, Durairaja A, Endres T, Lessmann V, Fendt M. BDNF haploinsufficiency induces behavioral endophenotypes of schizophrenia in male mice that are rescued by enriched environment. *Translational Psychiatry*. 2021;11(1):233. <https://doi.org/10.1038/s41398-021-01365-z>.
24. Cases S, Saavedra A, Tyebji S, Giralto A, Alberch J, Pérez-Navarro E. Age-related changes in STriatal-Enriched protein tyrosine Phosphatase levels: Regulation by BDNF. *Molecular and Cellular Neuroscience*. 2018;86:41-9. <https://doi.org/10.1016/j.mcn.2017.11.003>.
25. Walker MP, LaFerla FM, Oddo SS, Brewer GJ. Reversible epigenetic histone modifications and Bdnf expression in neurons with aging and from a mouse model of Alzheimer's disease. *Age*. 2013;35:519-31. <https://doi.org/10.1007/s11357-011-9375-5>.
26. Ceballos-Picot I, Nicole A, Clément M, Bourre J-M, Sinet P-M. Age-related changes in antioxidant enzymes and lipid peroxidation in brains of control and transgenic mice overexpressing copper-zinc superoxide dismutase. *Mutation Research/DNAging*. 1992;275(3-6):281-93. [https://doi.org/10.1016/0921-8734\(92\)90032-k](https://doi.org/10.1016/0921-8734(92)90032-k).
27. Leyane TS, Jere SW, Houreld NN. Oxidative stress in ageing and chronic degenerative pathologies: molecular mechanisms involved in counteracting oxidative stress and chronic inflammation. *International journal of molecular sciences*. 2022;23(13):7273. <https://doi.org/10.3390/ijms23137273>.
28. Heurtaux T, Bouvier DS, Benani A, Helgueta Romero S, Frauenknecht KB, Mittelbronn M, et al. Normal and pathological NRF2 signalling in the central nervous system. *Antioxidants*. 2022;11(8):1426. <https://doi.org/10.3390/antiox11081426>.
29. Illes P, Rubini P, Ulrich H, Zhao Y, Tang Y. Regulation of microglial functions by purinergic mechanisms in the healthy and diseased CNS. *Cells*. 2020;9(5):1108. <https://doi.org/10.3390/cells9051108>.
30. Woo Y, Lim JS, Oh J, Lee JS, Kim J-S. Neuroprotective effects of euonymus alatus extract on scopolamine-induced memory deficits in mice. *Antioxidants*. 2020; 9 (5) :449. <https://doi.org/10.3390/antiox9050449>.
31. Cheedella HK, Silakabattini K, Siahmansur TJ, Ishaq BM. Evaluation Of Neuroprotective Activity In Scopolamine Induced Dementia In Wistar Rats By Using Various Pharmacological Equipment And Its Histopathology. *Journal of Survey in Fisheries Sciences*. 2023: 1299-307. <https://sifisheriesciences.com/index.php/journal/article/view/813/363>.
32. Lee JC, Park JH, Ahn JH, Park J, Kim IH, Cho JH, et al. Effects of chronic scopolamine treatment on cognitive impairment and neurofilament expression in the mouse hippocampus. *Molecular medicine reports*. 2018;17(1):1625-32. <https://doi.org/10.3892/mmr.2017.8082>.
33. Kim JH, Han Y-E, Oh S-J, Lee B, Kwon O, Choi CW, et al. Enhanced neuronal activity by suffruticosol A extracted from Paeonia lactiflora via partly BDNF signaling in scopolamine-induced memory-impaired mice. *Scientific Reports*. 2023;13(1):11731. <https://doi.org/10.1038/s41598-023-38773-8>.
34. Ban JY, Park HK, Kim SK. Effect of glycyrrhizic acid on scopolamine-induced cognitive impairment in mice. *International Neurology Journal*. 2020;24(Suppl 1):S48. <https://doi.org/10.5213/inj.2040154.077>.
35. Bae HJ, Sowndhararajan K, Park H-B, Kim S-Y, Kim S, Kim DH, et al. Danshensu attenuates scopolamine and amyloid- β -induced cognitive impairments through the activation of PKA-CREB signaling in mice. *Neurochemistry International*. 2019;131:104537. <https://doi.org/10.1016/j.neuint.2019.104537>.
36. Aykac A, Ozbeyli D, Uncu M, Ertas B, Kilinc O, Şen A, et al. Evaluation of the protective effect of Myrtus communis in scopolamine-induced Alzheimer model through cholinergic receptors. *Gene*. 2019; 689:194-201. <https://doi.org/10.1016/j.gene.2018.12.007>.
37. Anand A, Khurana N, Ali N, AlAsmari AF, Alharbi M, Waseem M, et al. Ameliorative effect of vanillin on scopolamine-induced dementia-like cognitive impairment in a mouse model. *Frontiers in Neuroscience*. 2022;16:1005972. <https://doi.org/10.3389/fnins.2022.1005972>.
38. Lee M-R, Yun B-S, Park S-Y, Ly S-Y, Kim S-N, Han B-H, et al. Anti-amnesic effect of Chong-Myung-Tang on scopolamine-induced memory impairments in mice. *Journal of ethnopharmacology*. 2010;132(1):70-4. <https://doi.org/10.1016/j.jep.2010.07.041>.
39. Sharma C, Kim SR. Linking oxidative stress and proteinopathy in Alzheimer's disease. *Antioxidants*. 2021;10(8):1231. <https://doi.org/10.3390/antiox10081231>

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الآثار الطويلة المدى للسكوبولامين على أنسجة المخ لدى الفئران

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الخلاصة

خلفية البحث: السكوبولامين هو دواء مضاد للكولين يعطل انتقال الكولين في الجهاز العصبي المركزي كما أنه يسبب تشوهات إدراكية وعلامات مرضية مشابهة لتلك التي تظهر في مرض الزهايمر. ولذلك، يتم استخدامه لتحريض مرض الزهايمر في النماذج الحيوانية.

الأهداف: كان الهدف من الدراسة الحالية هو دراسة آثار التحريض طويل المدى مع السكوبولامين على أنسجة المخ لدى الفئران.

طرق العمل: تم تقسيم سبعين فأراً بالغاً إلى مجموعتين متساويتين: المجموعة الأولى كانت مجموعة طبيعية ومراقبة تلقت الماء المقطر فقط. أما المجموعة الثانية فكانت مجموعة تحريض مرض الزهايمر حيث تلقت السكوبولامين داخل الصفاق (1 ملجم / كجم) لمدة 14 يوماً فقط بعد ذلك تم إعطاء الماء المقطر لمدة 6 أشهر التالية. تم عزل عشرة فئران من كل مجموعة في وقت الصفر، بعد أسبوعين من التحريض، وبعد 3 أشهر وبعد 6 أشهر، وتم إخضاعها للاختبارات السلوكية ثم تم تشريحها لتحديد العوامل البيوكيميائية (بما في ذلك عامل التغذية العصبية المشتق من الدماغ، وحالة مضادات الأكسدة الكلية، والمالونديالدهيد). و(الأميلويد β). وقد تم تحليل البيانات باستخدام اختبارات t، وANOVA. مع اعتبار جميع القيم المعبر عنها كقيمة متوسط \pm SD وقيمة $P < 0.05$ ذات دلالة إحصائية.

النتائج: أنتج السكوبولامين تغيرات نسجية مرضية في الدماغ مشابهة لتلك التي تحدث في مرض الزهايمر البشري. ومع ذلك، لم تنتج فروق ذات دلالة إحصائية أخرى في الاختبار السلوكي، والعلامات البيوكيميائية، خلال الفترة الإجمالية للدراسة.

الاستنتاجات: يُحدث السكوبولامين تغيرات في أنسجة المخ والتي تستمر لفترة طويلة ويمكن استخدامه لدراسة مرض الزهايمر على المدى الطويل.

الكلمات المفتاحية: الاجهاد التاكسدي؛ الوظيفة المعرفية؛ سكوبولامين؛ مرض الزهايمر؛ مضادات الأكسدة.