Molecular Detection and Genotyping of Human Papilloma Virus Infections in Iraqi Patients with Esophageal Carcinoma.

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Summary:

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Background: Molecular DNA hybridization has confirmed more than 120 different human papilloma virus (HPV) genotypes. A small group of them have high- risk oncogenic potential. Many studies have described an association of such high risk-HPV genotypes with a variety of esophageal benign tumors as well as malignant squamous cell carcinomas.

Patients and Methods: A total number of 90 tissue specimens were collected from 50 patients with esophageal squamous cell (SCC), adenocarcinoma (AC) and carcinoma in situ (CIS); 20 patients with squamous acanthosis (SA); and 20 individuals with apparently-healthy esophageal tissues (AHET). The molecular detection methods for HPV detection and genotyping were performed by in situ hybridization using cocktailed- and specific high- risk HPV DNA probes, respectively.

Results: The overall percentage of HPV in the total group of esophageal carcinoma was 20%. The percentage of HPV DNA in the subgroup of SCC and AC was 26.7% and 13.3%, respectively,. However, neither HPV DNA was detected in CIS subgroup nor in both control groups (SA and AHET). The overall genotyping results showed that HPV 18 constituted the majority of the detected high-risk oncogenic HPV genotypes, followed by HPV 16 then HPV 31/33.

Conclusions: Despite the low prevalence of HPV infection and rarity of invasive esophageal carcinoma in the general Iraqi population, the detection of high percentage of such high oncogenic risk- HPV genotypes in these carcinomas indicating for a relevant importance in esophageal carcinogenesis. **Key Words:** Esophageal Carcinoma; Human Papilloma Virus; In Situ Hybridization.

Introduction:

Esophageal cancer ranks the ninth among the most common cancers world wide and represents approximately 4% of the total cancers. It remains one of the leading causes of cancer mortality. It ranks the sixth most common of cancer-related deaths (1-3). An association of some viruses with the occurrence of esophageal cancer has been described, like: Herpes Simplex Virus (HSV), Cytomegalovirus, Epstein Barr virus (EBV) and Human Papilloma Virus (HPV) (4, 5). The papilloma viruses have a specific tropism for squamous epithelial cells and their full productive cycle is only supported in these cells (6). Update more Than 100 different HPV types have been described; of these, nearly 30 can infect the anogenital tract some of These HPVs are frequently associated with cancer and are considered high risk genotypes (for e.g. HPV type 16 and 18), whereas others give rise to warts and benign lesions and are considered as low and intermediate risk genotypes (for e.g. HPV type 6, 11, 31, 33 and 35) (4, 5, 7).

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The association of HPV infections in squamous cell precancerous lesions of the uterine cervix has been well established since the late 1970 (8) Oncogenic HPV types are regarded as the most important etiological factor of cervical SCC.(9, 10, 11). The early 1980s have witnessed the rapid expansion of HPV research from genital tract to cover the other squamous cell epithelia, thus widening the scope of HPV-associated human tumors to involve the oral cavity, pharynx, larynx, upper airway, and esophagus (8, 10, and 12). The squamous cell lining of the oral mucosa is in direct continuity with the esophagus and the first descriptions on HPV lesions in the oral mucosa were slightly preceded by reports suggested by Syrjanen in 1982 that there is an association between HPV with both benign and malignant squamous cell lesions of the esophagus (13). Hille and co-workers were the first who used IHC in 1986 for the demonstration of HPV antigens in esophageal SCC (14). Currently, PCR and DNA hybridization are used for HPV genotyping (15). Following these pioneering observations, HPV research has resulted in rapidly expanding literature on both benign and malignant esophageal lesions in different geographical regions (16, 17, and 18). In addition, the evidence accumulated during the past 20 years is strongly suggestive of a

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causal role for HPV in esophageal carcinogenesis (7, 19, and 20). However, data concerning the involvement of HPV in esophageal cancers are still controversial and the exact mechanism by which HPV contributes to carcinogenesis has yet to be elucidated in the context of the esophagus (21, 22, and 23). This type of cancer is rare in Iraq, where is represent 0.7% among other cancers (24). The present study represents the first report in Iraq that designed to study the prevalence as well as the genotypes of HPV infection in Iraqi patients with esophageal cancers to elucidate a possible etiological role for HPV in esophageal carcinogenesis.

Materials and Methods:

Subjects: This study was designed as a retrospective research. A total number of (90) formalin-fixed, paraffin embedded esophageal tissue blocks were collected during the period from August 2004 to March 2005. These patients had undergone esophageal biopsy. They were collected from the archives of histopathology laboratories of different general hospitals, including Specialized Surgeries Hospital laboratories in the Medical City, AL-Yarmook Teaching Hospital Laboratories, Al-Kadhemiya Teaching Hospital Laboratories, Gastroenterology and Liver Diseases Teaching Hospital Laboratories, Al-Kendi Teaching Hospital Laboratories, Baquba Public Hospital Laboratories, Azadi (Kirkuk)Public Hospital, as well as from many private laboratories, including Dr.Raji Al-Hadethi private laboratory, Dr.Loui AL-Khuri private laboratory, Dr.Nawal Alash private laboratory and Dr.Faris Khairo private laboratory. Fifty esophageal malignant samples has been collected from 30 patients with squamous cell carcinoma (SCC) and 15 specimens from adenocarcinoma (AC) and 5 from those with carcinoma in situ .In addition, 20 tissue samples from squamous acanthosis cases were included, as a related esophageal disease control group, as well as 20 esophageal tissue samples with normal histological appearance (i .e without any significant pathological changes) were also included, as healthy control group for this study. The diagnosis of these tissue blocks were based on the obtained histopathological records attached to each esophageal biopsy in the relevant hospital files that accompanied these specimens. But following trimming process of these tissue blocks, a confirmatory histopathological re-examination of each obtained tissue blocks was done by further consultant histopathologist.

Methodology: Molecular detection and genotyping of HPV DNA in those tissue blocks was performed by a recent generation of in situ hybridization (ISH), using a specific biotinylated DNA probes for high oncogenic-risk HPV genotypes including 16, 18, 31/33.

Tissue Sectioning and Slide Preparation: At the histopathological department of Teaching laboratories

/ Medical City, formalin-fixed paraffin-embedded blocks from each esophageal biopsy was subjected to cut as serial thin sections of (4-5µm) thickness and were sticked on charge slides In order to prevent carryover DNA contaminations from one tissue sample to another, only one disposable cutting knife, which was specified for each tissue block, was used and then each section was sticked on a single charged slide. The 1st and 2nd tissue slides were specified for hematoxyline and eosin staining whereas many subsequent 4-5µm thickness-paraffinized tissue sections were specified for the following procedures of in situ hybridization. As positive and negative HPV controls, it was feasible to include such tissue-containing charged slides in each experiment by using cervical tissue blocks, proved by PCR to have both cocktailed and high riskoncogenic HPV genotypes, as a positive controls, as well as negative control from those apparently healthy cervical tissues, that were also proved by PCR technique to be negative for HPV.

In Situ Hibridization for Detection Cocktailed (Generic) – HPV Genotypes: The detailed instructions of the processes for performing in situ hybridization method for detection and genotyping HPV in this study were done according to the manufacturing company (Dako corporation Co./ Denmark) (25).

In Situ Hybridization of HPV Genotype 16, 18, 31/33 Biotinylated and Probes: The same procedures for in situ hybridization for detection of cocktailed probes of HPV (Dako cytomation Code No. K0601) were followed, except for the step of the temperature for stringent washing of these steps was changed to 58°C for 20-40 minutes. Assessment of the Result: Within 2 hours, the obtained result was assessed by examining the processed slides under light microscope; a deposition of a soluble blue purple product at the sites of hybridization of the probes to their targets is a positive indicator for the presence of the questioned group of HPV. Statistical Analysis: T test, and ANOVA test, and Qui square were applied for statistical examination of all results obtained in our research.

Results:

1. Detection in the total group of esophageal carcinoma was 20%, while its percentage in the SCC group was 26.7% and in the AC group was 13.3%. No HPV DNA was detected in both squamous acanthosis and healthy control group (Table 1 & Figure 1).

Table1: Percentage of cocktailed HPV DNADetection in esophageal carcinoma

Study Groups	No. of Tested Cases	Cocktailed HPV DNA Detection Via In Situ Hybridization Positive Negative			
		No.	%	No.	%
Carcinoma In Situ	5	0	0.0	5	100.0
Squamous Cell Carcinoma	30	8	26.7	22	73.3
Adenocarcinoma	15	2	13.3	13	86.7
Squamous Acanthosis	20	0	0.0	20	100.0
Healthy Control	20	0	0.0	20	100.0



Figure1: Positive in situ hybridization reaction showing HPV-DNA (Generic) within the cells of tissue blocks from patients with esophageal cancer. BCIP/NBT-chromogen stained & counter stained by nuclear fast red (X100)

2. Sex Distribution in HPV-Associated Esophageal Carcinoma:

None of the cases with carcinoma in situ had showed cocktailed / generic HPV DNA-ISH reaction, whereas HPV DNA was detected in 6 males (20%) out of 30 SCC and in 2 females (6.6%) out of 30 SCC. In those cases with AC, HPV DNA was detected in only 2 males (13.3%) out of 15 AC specimens (table 2).

 Table 2: The prevalence of generic HPV DNA in esophageal carcinoma according to sex

Study Groups		Male			Female				
	Tota	HPV positive		HPV negative		HPV positive		HPV negative	
	1	No	%	No	%	No	%	No	%
Carcinoma In Situ	5	0	0.0	3	60. 0	0	0. 0	2	40. 0
Squamous Cell Carcinoma	30	6	20. 0	14	46. 7	2	6. 6	8	26. 7
Adenocarcinom a	15	2	13. 3	6	40. 0	0	0. 0	7	46. 7
Squamous Acanthosis	20	0	0.0	12	60. 0	0	0. 0	8	40. 0
Healthy Control	20	0	0.0	14	70. 0	0	0. 0	6	30. 0

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3. Genotyping Results of HPV- Positive Esophageal Carcinoma:

Three different HPV genotypes were found in cases with esophageal carcinoma of the present study. The genotype HPV16 was detected in 5 out of 8 (62.5%) of HPV-positive SCC tissue blocks. However, no HPV 16 was detected in any AC specimens (Table3 & Figure 2).

 Table3: Frequency of HPV- genotype 16 detection

 in esophageal carcinoma

Study Groups	No. of HPV	In Situ Hybridization for HPV genotype 16 Detection				
	-Positive- tested cases	Positive	9	Negative		
		No.	%	No.	%	
Squamous Cell Carcinoma	8	5	62.5	3	37.5	
Adenocarcinoma	2	0	0.0	2	100.0	
Total Esophageal Carcinoma Group	10	5	50.0	5	50.0	



Figure 2: Positive in situ hybridization reaction showing HPV genotype 16 DNA within the cells of tissue blocks from patients with esophageal cancer. BCIP/NBT-chromogen stained & counter stained by nuclear fast red (X200).

On the other hand, HPV18 was detected in half (50%) of the HPV-positive SCC specimens, whereas all adenocarcinoma specimens were positive for HPV18 (Table 4).

Table4: Frequency of HPV- genotype 18 in esophageal carcinoma.

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Study Groups	No. of HPV- Positive Tested	In Situ Hybridization for HPV genotype 18 Detection							
	Cases	Positiv	'e	Negative					
		No.	%	No.	%				
Squamous Cell Carcinoma	8	4	50.0	4	50.0				
Adenocarcinoma	2	2	100.0	0	0.0				
Total Esophageal Carcinoma Group	10	6	60.0	4	40.0				

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The HPV31/33 in the current study was detected in 2 out of 8 (25%) of SCC cases and in half (50%) of adenocarcinoma cases (table 5).

Table 5: Frequency of HPV genotype 31/33 inesophageal carcinoma

Study Groups	No. of HPV-	In Situ Hybridization for HPV genotype 31/33 Detection					
	Positive	Positive Negative			ive		
	Tested Cases	No.	%	No.	%		
Squamous Cell	8	2	25.0	6	75.0		
Carcinoma							
Adenocarcinoma	2	1	50.0	1	50.0		
Total Esophageal	10	3	30.0	7	70.0		
Carcinoma Group							

Lastly, it was found in this study tissue blocks with mixed- HPV genotypes infections in 4 out of 8 (50%) of HPV-positive esophageal carcinoma cases (table 6).

Table 6: Frequency of mixed HPV genotypesDetection in esophageal carcinoma

Detection in esophagear caremonia									
Study	No.	Mi	xed	HPV	HPV	HPV	HPV		
Groups	of	HPV		Geno-	Geno-	Gen-	Gen-		
_	HPV			type	type	otype	otype		
	-			16+18	16+	18+	16+18		
	positive				31/33	31/33	+31/33		
	Tested	No.	%	Freq-	Freq-	Freq-	Freq-		
	blocks			uency	uency	uency	uency		
Squamous	8	4	50.0	1	1	1	1		
Cell									
Carcinoma									
Adeno-	2	1	50.0	0	0	1	0		
carcinoma									

Discussion:

Results of HPV Detection in Esophageal Carcinoma: Data concerning the involvement of HPV in esophageal cancers are controversial (23). This type of cancer is rare in Iraq. It represents 0.7% among other cancers. The present study represents the first report regarding the prevalence of HPV infection in Iraqi patients with esophageal cancer to elucidate a possible etiological role for HPV in esophageal carcinogenesis. In the current study, the esophageal tissue samples, which were collected from histopathology laboratories of different general hospitals as well as many private laboratories, are including the following specimens:-

A. Five carcinoma in situ samples; none of them were positive for HPV using recent generation of ISH techniques. This could be related to the small number of biopsies that were studied as well as to the very tiny size of these biopsies that were endoscopically biopsied (i.e. they are tiny pieces) and are not like those surgically resected specimens and thus could lead to a missing HPV-positive esophageal precancerous cases. In consistence with the present results, only one study that examined both endoscopic biopsies and surgically-resected specimens and found a lower incidence of HPV DNA in endoscopic biopsies (26). However, other studies were able to demonstrate a high incidence of HPV DNA but they had relied exclusively on large numbers of biopsies (27).

B. The HPV DNA are detected in 8 out of 30 (26.7 %) of invasive SCC cases whereas only 2 out of 15 (13.3%) of invasive AC cases are HPV- positive. In SCC, the present HPV- positive results are as double higher as those positive results of HPV in AC cases. This could be related to the fact that HPV are epitheliotropic viruses which require squamous epithelial tissue as micro- environment for their replication and to continue their life cycle (28).On reviewing and discussing the relationship of HPV to esophageal AC, it was noticed that only very limited researches in this field were present and only few researchers had directed their efforts in this field; one of them, in Turkey, who scored HPV in 4 out of 10 (40%) of AC cases by using PCR technique (3). Another study in China, had also detected the HPV score but in 22 out of 62 (35.5%) of the AC of the gastric corpus by using ISH technique which is a solid tumor similar to AC of the esophagus (29). The frequency of HPV infection, reported in the present study, is placing our country among countries in the moderate risk region especially when we compared the detected prevalence with those in high and low risk regions of HPV infection. In high- risk regions (such as China and South Africa) it had reported HPV prevalence that ranged from 40 to 71%) (30). Chang and co-workers(16) reported the presence of HPV in 43% of Chinese esophageal carcinoma cases whereas HPV-DNA was detected in 52%, 65% and 71% of esophageal carcinoma cases from South Africa (30). Moreover, lower frequencies (2% and 6.7%) were reported in western countries and North America, respectively (26). In these low- risk regions, most studies have failed to find any association between HPV and esophageal cancer (31). The present results are ranked between the results of very high risk regions of HPV infection and the other extremity (zero% of HPV detection). The examples of studies from these very high risk regions are those of Chen et al (32) who had demonstrated HPV positivity in 60% of SCC cases from high risk areas of China and study of Williamson et al (33) who also reported HPV DNA positivity in 71% of esophageal cancer tissue. While reports from Japan, North Europe and United States show little or no evidence of HPV in esophageal SCC (34, 35). A potential shortcoming of this study, as well as other retrospective studies, are their limited ability to find out a clear-cut explanation for the association between HPV and those cases of esophageal cancer, whether HPV has a hit-and-run mechanism in this anatomic site or not. By analogy, bovine papilloma virus-4(BPV-4) causes diffuse alimentary tract papillomatosis in cattle. This infection is usually a self-limited, unless the cattle were

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ingesting broken fern. It was observed that the bracken fern contained both immunosuppressant and a DNA agent (called quercetin). damaging Those immunosuppressed cattle are unable to clear the BPV-4-infected cell and thereby promote tumor progression (36). Despite its presence in benign papilloma, the BPV is rarely found in malignant tumors and this notice is suggesting an important role for the virus in tumor initiation and promotion but not maintenance of the malignant phenotype (12). However both the absence of HPV in benign esophageal lesions and the lack of an association between HPV L1 seroactivity and esophageal cancer risk argue against this possibility in western cultures, here, further case series and case-control studies, especially in high-incidence areas, such as Africa and China were warranted. Such studies should include detailed evaluations of human dietary or medical exposures that may be analogous to broken fern in cattle (12). No infection was revealed by ISH techniques in both squamous acanthosis group and healthy control group in this study. Moreover, marked variations in the prevalence of esophageal HPV infections have been reported by different authors. These differences might be, in part, due to the differences in the sensitivity and specificity of the methods used. Furthermore, most researchers who made relation between HPV infection and esophageal cancer had especially studied the relationship between HPV and esophageal SCC that are, in one hand, based on the observation of characteristic histopathological findings (koilocytic changes), suggesting the presence of HPV in benign esophageal epithelia and malignant esophageal tumors and, in the other hand, are based on the morphological similarities between esophageal SCC and cervical SCC (13). The consumption of tobacco and alcohol could explain more than 90% of cases of esophageal SCC (37). Also health registries had indicated that the consumption of tobacco by Iraqi people is very high which in turn may enhance the possibility of SCC of the esophagus (24).In addition, in some areas with a high risk of esophageal cancer, like China and Latin America, the consumption of very hot beverages (hot mate or boiling water) and the consumption of fermented fish have been identified as probably carcinogenic factors for esophagus (38).It is worthy to mention that Iraqi people have a similar disposition to those in southern east of Asia in respect to same habits of eating spicy foods as well as consumption hot beverages (like tea) which were well recognized as one of the risk factors.

Genotype Results of HPV-Positive Esophageal Carcinoma: The mechanisms through which HPV16 and HPV18 can induce epithelial neoplasia have been extensively studied (39, 40). Some of their viral proteins, notably E6 and E7 are oncoprotiens that immortalize various human cell types, inactivate host protein (such as p53 or Rbp) and induce mutations in the host cell DNA (41). Since the most prevalent HPV genotypes in different studies were related to the class of high oncogenic types (i.e. HPV16 and HPV18), most of these studies had suggested a possible etiological role for HPV in esophageal cancer .The other reported HPV genotypes were supposed to represent a lower oncogenic HPV risk types, these were HPV6, HPV11, HPV31 and HPV 33 (30). On comparing the results of the present study with 2 separate studies in Turkey and Iran, which had also reported the presence of HPV16 and HPV18 in both esophageal SCC and AC by PCR technique; in Turkey, the researchers had detected HPV16 and /or HPV18 in 10 out of 30 (33.3%) of SCC samples and 1 out of 10 (10%) of AC samples (3) .In the Iranian study, HPV16 and HPV18 have been found in 5 out of 38 (13.2%) and in 3 out of 38 (7.9%) of esophageal SCC cases, respectively (23). The differences noticed in these percentages of HPV16 and HPV18 between these studies when we compared them to our results they may be due to different techniques used in each study. Moreover, in those studies there may be other HPV genotypes in the examined materials but they were not detected because they could be out of the range of the ability of detection level of the used HPV probes. The results of this study are similar to the abroad one where it was observed that this disease is common in men more than women. SCC had noticed to occur most commonly in African American men compared with Caucasian men, African American women, and Caucasian women. Although SCC occurs more commonly in men than women in areas of low incidence, however, high incidence areas did not show such strong sex difference (42). Esophageal AC is the fastest rising malignancy among Caucasian men. Men were 6-8 fold more likely than women to suffer from esophageal AC and Caucasians were 3-4fold more likely than African Americans (43). Also the effect of such sex differences was noticed when HPV genotyping of HPV positive cases were done in our study especially HPV genotypes 16, 18 and 31/33 where in this respect, all of them were higher in males more than females. These results were consistent to the results of (42). In this study, positive HPV-results were detected in old aged patients. For SCC, the age range was between (40-80 years) for males and for females was (65-75 year). However, there were no HPV-positive results in females. So far, our results are in good agreement with the results of Yang and Davis who demonstrated an age-related increase risk for the development of SCC and AC (31). The incidence for each of these cancers is extremely low under the age of 40 years. After the age of 40, an increased incidence had been observed with each decade of life (31, 44).

Conclusion:

1. Despite a low prevalence of HPV infection and the rarity of invasive esophageal carcinoma in the general Iraqi population, the detection of differed oncogenic HPV genotypes in that esophageal cancer could have a relevant importance, among other risk factors, in the process of its carcinogenesis.

2. The very lower percentage of HPV DNA in preinvasive esophageal neoplastic lesions in the present study (than previous studies elsewhere) could be in part, due to other HPV genotypes (but not included with the range of cocktailed-genotypes probes used for in situ hybridization in this study) or due to different other agents that have oncogenic potential or have some role in the carcinogenesis of HPV-negative esophageal carcinoma. In addition, carcinoma in situ group that showed a negative result for generic HPV infection could also be a reflection that most of these HPV infections are transient and did not persist or might be obstacle by the very low number of specimens or are not present in these very tiny pieces (endoscopic biopsies) used in these experiments.

References:

1 .Blot WJ, Devesa SS, Fraumeni JF: Continuing climb in rates of esophageal adenocarcinoma: an update. JAMA 1993; 270:1320.

2. Haboubi NY, Geboes K., Shepherd NA, Talbot IC: Gastrointestinal polyps. Greenwich medical media limited. San Francisco. 1st edition: 3-19, 2002.

3. Kiki I, Gündoădu M., Polat F, Gündoău C: Detection of human papilloma virus infection in esophageal carcinomas by the histopathological method and polymerase chain reaction technique. Turk J Med Sci 2002; 32:223-230.

4. Oriel JD: Historical Overview. In: Human Papillomavirus in Dermatovenereology. Gross, G., Von Krogh, G: (editors). Boca Raton, CRC press inc. 1:7-12, 1997.

5. Herrington CS, Graham D, Southern SA, Bramdev A., chetty R: Loss of Retinoblastoma protein expression is frequent in small cell neuroendocrine carcinoma of the cervix and is unrelated to HPV type. Hum Pathol 1999; 30(8): 906-910.

6. Giroglou T, Florin L, Schafer F, Streeck RE, Sapp M: Human Papillomavirus infection requires cell surface Heparan sulfate, J Virol 2001; 75(3): 1565-1570.

7. Mathers CD, Shibuyak R, Boschi—pinto C : Global and regional estimates of cancer mortality and incidence by site: I. Application of regional cancer survival model to estimate cancer mortality distribution by site. BMC Cancer 2002; 2(1): 36.

8. Syrjänen K, Gissmann L, Koss LG, et al: Papillomaviruses and human disease. Berlin: Spring Verlag, 1987.

9. IARC Monographs on the evaluation of carcinogenic risks to humans. Papillomaviruses, Lyon: IARC. Vol. 64, 1995.

10. Syrjänen K, Syrjänen S. Papillomaviruses infections in human pathology. New York: Wiley & Sons, 2000.

11. Bosch XF, Lorincz A, Muñoz N: The causal relation between human papillomavirus and cervical cancer. J Clin Pathol 2002, 55:244-65.

12. Gillison ML, Shah KV : Role of mucosal human papillomavirus in nongenital cancers. J Natl Cancer Inst Monogr 2003; 31:57-65.

13. Syrjänen K: Histological changes identical to those of condylomatous lesions found in esophageal squamous cell carcinomas. Arch Geschwulstforsch 1982; 52:283-92.

14. Hille JJ, Margolius KA, Markowitz S, et al: Human papillomavirus infection related to esophageal carcinoma in black South Africans. A preliminary study. S Afr Med J 1986; 69:417-20.

15. Poseschla, EM, Wong-Staal F (1997): Etiology of cancer: Viruses. In: Cancer Principles and Practice of Oncology, 5th edition.DeVita, V.T.; Hellman, S.; and Rosenberg, S.A (editors). Lippincott-Raven Publishers Philadelphia Chapter 8; 153-184, 1997.

16. Chang F, Syrjanen S, Shen Q, et al. Human papillomavirus involvement in esophageal carcinogenesis in the high-incidence area of China: A study of 700cases by screening and type-specific in situ hybridization. Scand J Gastroenterol 2000; 35: 123-30.

17. Syrjänen K, Syrjänen S. Papillomaviruses infections in human pathology. New York: Wiley & Sons, 2000.

18. Tripodi S, Chang F, Syrjänen S, et al. Quantitative image analysis of esophageal squamous cell carcinoma from the high-incidence area of China, with special reference to tumor progression and papillomavirus (HPV) involvement. Anticancer Res 2000; 20:3855-62.

19. Poljak M, Cerar A, Seme K. Human papillomavirus infection in esophageal carcinomas: a study of 121 lesions using multiple broad-spectrum polymerase chain reactions and literature review. Hum Pathol 1998; 29: 266-71.

20. Zur-Hausen H. Papillomaviruses in human cancers. Proc Assoc Am Physicians, 1999; 111:581-587.

21. Yasuda M, Kuwano H, and Watanabe M, et al.: P53 expression in squamous dysplasia associated with carcinoma of the esophagus: evidence for field carcinogenesis. Br J Cancer 2000; 83: 1033-1038.

22. Breton J, Sichel F, Abbas A, et al: Simultaneous use of DGGE and DHPLC to screen TP53 mutations in cancers of the esophagus and cardia from European high incidence area (Lower Normandy, France). Mutagenesis 2003; 18:229-306.

23. Farhadi M., Tahmasebi Z, Merat S., et al: Human papilloma virus in squamous cell carcinoma of esophageal in high-risk population. J of Gastroenterol, 2005; 11(8): 1200-1203.

24. Iraqi Cancer Board: Results of Iraqi Cancer Registry 1995-1997. Ministry of health (editor). Baghdad. Iraq. P. 11-13, 34, and 32-39, 1999.

J Fac Med Baghdad

25. Al-jewari MMM, Mohammed-Ali SH, Al-azzawi MKK: Genotyping of Human Papilloma Virus infections and phenotyping of tumor infiltrating lymphocytes in Iraqi patients with uterine cervical neoplasia. The Iraqi Postgraduate Medical Journal 2007; 6 (4):362-373.

26. De Villiers E M, Lavergne D, Chang F, et al: An interlaboratory study to determine the presence of human papillomavirus DNA in esophageal carcinoma from China. Int J Cancer 1999; 81: 225-8.

27. Woo Y J, Yoon HK: In situ hybridization study on human papillomavirus DNA expression in benign and malignant squamous lesions of the esophagus. Journal of Korean Medical Science 1996; 11:467-73.

28. Bodaghi S, Wood LV, Roby G, et al: Could human papilloma virus spread through blood?. J Clin Microbiol, 2005; 43(11):5428-5434.

29. Zhang X., Lu ZM, Li JY, et al: Detection of human papilloma virus 16 E 6 mRNA in carcinomas of upper digestive tract J.pub Med, 2003; 83(21): 1910-1914.

30. Bahnnassy AA, Zekri AN, Abdalah S, et al: Human papilloma virus infection in Eygptian esophageal carcinoma: Correlation with p53, $p21^{waf}$, mdm2, C-erb B₂ and impact on survival. Pathology international, 2005; 55:53-62.

31. Syrjanen K: HPV infections and esophageal cancer. J Clin Pathol 2002; 55:721-8.

32. Chen B, Yin H, Dhurandhar N. Detection of Human Papillomavirus DNA in Esophageal Squamous cell Carcinomas by the Polymerase China Reaction Using General Consensus Primers. Hum. Pathol 1994; 25: 920-3.

33. Williamson AL, Jaskiesicz K, Gunning A. The Detection of Human Papillomavirus in esophageal Lesions. Anticancer Research 1991; 11: 263-6.

34. Kamath A, Wu T, Heitmiller R, et al: Investigation of the association of esophageal carcinoma with human papillomaviruses Dis Esoph 2000; 13:122-24.

35. Van Doornum GJ, Korse CM, Buning-Kager JC, et al. Reactivity to human papillomavirus type 16 L1

virus-like particles in sera from patients with genital cancer and patients with carcinomas at five different extragenital sites. Brit J Cancer 2003:88: 1095-100.

36. Beniston RG, Morgan IM, O'Brien V, Campo MS: Quercetin, E7 and p53 in papillomavirus oncogenic cell transformation. Carcinogenesis 2001; 22:1069-76.

37.Brown LM, Hoover R, Silverman D, et al: Excess incidence of squamous cell esophageal cancer among US Black men: Role of social class and other risk factors. Am J Epidemiol 2001; 153: 114-122.

38. Ke L, Yu P, Zhang ZX.: Novel epidemiologic evidence for the association between fermented fish sauce and esophageal cancer in south China. Int J Cancer 2002; 99:424-6.

39. Chen Hheng SB, Chew EC. Human papillomavirus 16 E6 is associated with the nuclear matrix of esophageal carcinoma cells. World J Gastroenterol 2001; 7: 788-791.

40. Munoz N, Bosch FX, de Sanjose S, et al: Epidemiologic classification of human papillomavirus types associated with cervical cancer. N Engl Med 2003; 348: 518-527.

41. Caldeira S, de Villiers EM, Tommasino M. Human papillomavirus E7 proteins stimulate proliferation independently of their ability to associate with retinoblastoma protein. Oncogene 2000; 19: 821-826. 42. National Cancer Institute, DCCPS, Surveillance Research program cancer Statistics Branch SEER

Research program, cancer Statistics Branch. SEER Program Public Use Data, 1973-1999 November 2001.

43. El-Serag HB. The epidemic of esophageal adenocarcinoma. Hematol Oncol Clin North Am 2003; 17:421-440.

44. Yang PC, Davis S. Incidence of the Cancer of the EsophagusintheUSbyHistologictype.Cancer1988; 61:612-17.